

This product is for research use only (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS20 4432

Anti-Fd3 | Ferredoxin 3

Product information

Immunogen Purified full length, tag cleaved, recombinant maize Fd3, UniProt: P27788

Host Rabbit

Clonality Polyclonal

Purity Total IgG. Protein A purified in PBS, 50% glycerol. Filter sterilized.

Format Liquid at 1 mg/ml.

Quantity 100 μg

Storage

Store at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution 1: 2000 - 1: 10 000 (WB)

Expected | apparent

16 kDa

Confirmed reactivity | Arabidopsis thaliana, Zea mays

Predicted reactivity Brachypodium distachyon, Dichanthelium oligosanthes, Hordeum vulgare, Oryza sativa, Panicum hallii, Saccharum sp.

Setaria italica Species of your interest not listed? Contact us

Not reactive in No confirmed exceptions from predicted reactivity are currently known

Additional information Antibody reacts weakly with other ferredoxins: Arabidopsis thaliana and Zea mays Fd2 and Zea mays Fd6.

Selected references Hanke and Hase (2008). Variable Photosynthetic Roles of Two Leaf-Type Ferredoxins in Arabidopsis, as Revealed by

RNA Interference. Photochem Photobiol. 84(6):1302-9. doi: 10.1111/j.1751-1097.2008.00411.x.

Hanke et al. (2003). A Post Genomic Characterization of Arabidopsis Ferredoxins. Plant Physiol. 134(1):255-64. doi:

10.1104/pp.103.032755.

Matsumura et al. (1997). A Nitrate-Inducible Ferredoxin in Maize Roots. Genomic Organization and Differential Expression of Two Nonphotosynthetic Ferredoxin Isoproteins. Plant Physiol. 114(2):653-60. doi: 10.1104/pp.114.2.653.



Fd3 purified from leaves of Zea mays, 50 ng was denatured with 4X SDS buffer at 95°C for 5 min. Samples were separated on 10% SDS-PAGE and blotted 1h to PVDF membrane. Blot was blocked with 3 % skim milk/TBS-T, 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 2000 in TBS-T for 1h/RT. The antibody solution was decanted and the blot was washed 4 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1h/RT with agitation. The blot was washed as above and developed with a chemiluminescent detection reagent, following manufacture's recommendation.



This product is for research use only (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Total leaf extract of Arabidopsis thaliaa (1) and Zea mays (2), 10 µg denatured with 4X SDS buffer at 95 °C for 5 min. Samples were separated on 10% SDS-PAGE and blotted 1h to PVDF membrane. Blot was blocked with 3 % skim milk/TBS-T, 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 10 000 in TBS-T for 1h/RT. The antibody solution was decanted and the blot was washed 4 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1h/RT with agitation. The blot was washed as above and developed with a chemiluminescent detection reagent, following manufacture's recommendation.

The antibody is also recognizing root-type ferredoxins expressed in leaves.



Different RNAi (lane 1-7) expressed in T1 *Arabidopsis thaliana* plants and wild type without any RNAi epxression. were freshly extracted with 2x SDS-sample buffer (+ 2ME) for SDS-PAGE. For IP, 150mM NaCL, 1% Triton X-100, 50 mM Tris-HCl (pH 8.0) and denatured with 4X SDS buffer at 95°C for 5 min. Samples were separated on 10% SDS-PAGE and blotted 1h to PVDF membrane. Blot was blocked with 3 % skim milk/TBS-T, 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 5000 in TBS-T for 1h/RT. The antibody solution was decanted and the blot was washed 4 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1h/RT with agitation. The blot was washed as above and developed with a chemiluminescent detection reagent, following manufacture's recommendation.